EFFECT OF ECHINACEA PURPUREA AND CLOVE (SYZYGIUM AROMATICUM) AQUEOUS EXTRACT ON IMMUNE DEFICIENCY INDUCED BY CISPLATIN IN RATS

Asmaa M. Abd El-Hafez^{*} ; Omnia G. Refaat ; H.G. El-Masry and Alaa O. Aboraya

Nutrition and Food Science Department, Faculty of Home Economics, Helwan University, Cairo, Egypt

*E-mail-asmaa892006@gmail.com

ABSTRACT

The present study was aimed to investigate the effect of Echinacea purpurea and clove (Syzygium aromaticum) aqueous extract on raising the immune response in rats with immune deficiency diseases. Forty-eight male albino rats were randomly divided into 8 equal groups (n=6). Group 1 was negative group, whereas the other 7 groups injected by cisplatin. Group 2 kept as the immunotoxic control group (+ve group). Groups 3 and 4 received 1 and 2 ml of *Echinacea* extract, whereas 1 and 2 ml of clove extract were given to groups 5 and 6, respectively. Groups 7 and 8 received 1 and 2 ml of Echinacea and clove extract mixture, respectively, for 8 weeks. Results of body weight, spleen weight, serum immunogolobulin G, immunogolobulin M (IgM), total protein, albumin, globulin, neutrophil, lymphocyte, monocytes and eosinophil significantly decreased. Whereas leukocyte significantly increased by cisplatin in positive control group as compared to the negative control group. On the other hand, administration of Echinacea and clove extracts and their combination attenuated these adverse effects and markedly ameliorated biochemical alterations that caused by cisplatin administration. In conclusion, Echinacea and clove extracts stimulate the immune system of rats with cisplatin-induced suppressed immunity. This study recommended increasing the consumption of the *Echinacea* and clove in the diet, this may be raising the immune response in patient with immune disorders.

Key Words: Echinacea purpurea, Clove, Cisplatin, Immune System, Rats.

INTRODUCTION

Cisplatin (also known as cisplatinum or *cis*diamminedichloroplatinum) is the first Food and Drug Administration, it is a platinum-containing compound which inhibits synthesis of RNA, DNA and protein in cells. Cisplatin is one of the most effective anticancer drugs used for the treatment of various oncologic diseases, including testicular, cervical, ovarian, mammary, head and neck,

esophageal, lung and brain cancers (Dasari and Tchounwou, 2014 and Zhou et al., 2022). Several adverse effects including myelosuppression, hepatotoxicity, nephrotoxicity and immunotoxicity are the result of cisplatin administration (Hasaaan et al., 2010). The toxic effects of cisplatin are attributed to several factors, such as peroxidation of the cell membrane, DNA damage, mitochondrial dysfunction, inhibition of protein synthesis, and ability to affect host immune response (Zhu et al., 2017). Immunotoxicity could be the result of direct or indirect action of a chemical on the immune system, causing a suppression or activation of the immune response (Tumeh et al., 2014). Compromised immune response can result in suppression of host resistance to infectious agents as well as tumor cells. Most anticancer drugs are found to suppress hematopoiesis in bone marrow and cause myelosuppression and lymphocytopenia, resulting in reduction or inhibition of lymphocytic responses. Cisplatin has a cytotoxic effect on immune cells when they are rapidly dividing.

Several plants and herbs are being used as immunomodulators (Nagoba and Davane, 2018; Zhang *et al.*, 2020). *Echinacea* is a plant genus within the family of Asteraceae (compositae) and is comprised of 11 taxa of herbaceous and flowering plants (Sharifi-Rad *et al.*, 2018). It is an indigenous medicine of the native American Indians and Europeans with multiple biological activities, such as anti-inflammation, anti-oxidation and immunomodulation effects (Chiou *et al.*, 2017 and Khattab *et al.*, 2019). *Echinacea purpurea* contains active ingredients of carbohydrate, glycoside, alkaloide, alkylamide and polyacetylene (Lalone *et al.*, 2007).

Clove, *Syzygium aromaticum*, is an aromatic medical plant of the family Myrtaceae. It is commonly applied as a natural additive in the food industry, antiseptic against infectious diseases, and local anesthetic in dentistry (**Cortés-Rojas** *et al.*, **2014**). In addition to its antimicrobial, anti-fungal, and anti-viral properties, clove possesses anti-inflammatory properties (**Chaieb** *et al.*, **2007**). Eugenol, eugenyl acetate, carvacrol, tanene, and thymol were detected as major constituents of the clove (**Amelia** *et al.*, **2017**). It has been reported that constituents of clove impart anti-oxidant activities and inhibit lipid peroxidation (**Dibazar** *et al.*, **2015**). Effects by clove or main constituents (like eugenol) on specific immune system components/mechanisms have only recently begun to be examined in detail (**Yogalakshmi** *et al.*, **2010**; **Bachiega** *et al.*, **2012 and Grespan** *et al.*, **2012**).

Therefore, this study was conducted to study the effect of aqueous extracts of *Echinacea purpurea* and Clove (*Syzygium aromaticum*) on raising the immune response in rats with immune deficiency diseases.

MATERIALS AND METHODS

Materials

Echinacea purpurea and Clove (*Syzygium aromaticum*) were obtained from Agriculture Research Center, Egypt. Cisplatin, casein, cellulose, choline chloride, D-L methionine, vitamin and mineral constituents were purchased from El-Gomhoriya Pharmaceutical Company, Cairo, Egypt. Starch, soy oil, and sucrose were obtained from the Egyptian local market. Forty-eight adult male rats (Sprague Dawley strain), weighing about 180±10 g b.wt. were obtained from the Laboratory Animal Colony, Agricultural Research Center, Giza, Egypt. **Methods**

1. Preparation of *Echinacea* and Clove Aqueous Extract:

Twenty-five gm of dried Echinacea was submerged in 100 ml of distilled water and allowed to soak overnight, then filtered to obtain a liquid extract. A known concentration of *Echinacea* aqueous extract was given orally by stomach tube.

The Clove flower buds were dried in the sun and ground to fine powder with the aid of an electric blender. Thereafter, 25 g of the milled clove powder was soaked in 100 ml distilled water, then filtered to prepare an aqueous extract (**Dibazar** *et al.*, **2015**). A known concentration of Clove aqueous extract was given orally by stomach tube.

2. Induction of Immune Deficiency Diseases:

Cisplatin-induced immune deficiency diseases in rats. Intraperitoneal injection of male albino rats with cisplatin (3.5 mg/kg) once every 3 days for consecutive 2 weeks (Wang *et al.*, 2013).

3. Diet Preparation and Experimental Design:

The basal diet was prepared according to AIN-93M diet (Reeves *et al.*, 1993). Forty-eight male albino rats were randomly divided into 8 equal groups (n=6). Group 1 was negative control group. Whereas the other 7 groups were injected by cisplatin. Group 2 was kept as the immunotoxic control group (+ve group). Groups 3 and 4 received 1 and 2 ml of *Echinacea* extract. Whereas 1 and 2 ml of clove extract were given to groups 5 and 6, respectively. Groups 7 and 8 received 1 and 2 ml of *Echinacea* and clove extract mixture (1:1), respectively, for 8 weeks. During the experiment period, the quantities of diet, which were

consumed and/or waste, were recorded every day. In addition, rat's weight was recorded weekly to determine body weight gain and feed efficiency ratio according to **Chapman** *et al.*, (1959).

4. Biochemical Analysis:

At the end of the experimental period (8 weeks), rats were fasted overnight before scarifying and blood samples were collected from each rat and centrifuged at 3000 rpm for 15 min to obtain the serum for biochemical analysis. Levels of leukocytes, neutrophil, lymphocyte, monocytes, eosinophil and basophil were estimated according to Ochei Kolharktar, (2008).Immunoglobulin and Μ (IgM) and immunoglobulin G (IgG) were measured according to Ziva and Pannall, (1984). Concentration of total protein was determined according to Burtis and Ashwood (1999), albumin and globulin were determined according to Young, (1995).

5. Statistical Analysis:

Results were expressed as the mean standard error \pm SE. Data were statistically analyzed for variance "ANOVA" test at P \leq (0.05) using SPSS statistical software, version 20 was used for these calculations according to Armitage and Berry, (1987).

RESULTS AND DISCUSSION

The results in **Table 1** show that intraperitoneal injection of cisplatin to rats caused a decrease in feed intake and a significant (P < 0.05) reduction in body weight gain (BWG%), feed efficiency ratio (FER) and spleen relative weight % when compared to the negative control group. Oral administration of *Echinacea* and Clove extracts and their mixture to rats inflicted with immune deficiency diseases (IDD) caused increasing in feed intake and a significant (P < 0.05) increases in FER, body weight and spleen weight as compared to the positive control group.

Lin *et al.*, (2018) demonstrated that cisplatin administration resulted in significantly decrease in feed intake, body weight and feed efficiency, as found in the present study. Immunotoxicity may parallel alterations in the weight of lymphoid organs (spleen) (Pearse *et al.*, 2009). The results in the present study were in the same line with EL-Sherbiny *et al.*, (2021) that the administration of *Echinacea* extract stimulated the increase in weight of spleen as well as body weight in rats with immune deficiency. Furthermore, Ali, (2008) reported that Echinacea has a positive effect on body weight gain after 4 weeks of treatment. Moreover, the obtained results agree with **Agbaje** *et al.*, (2009) who showed an improvement in body weight by clove in rats.

Table 1: Effect of Echinacea and Clove Extracts on Feed Intake (FI),Efficiency Ratio (FER), Body Weight Gain (BWG), andSpleen Relative Weight (SRW) of Rats with ImmuneDeficiency Diseases

Parameter	FI (g/d/rat)	FER	BWG%	SRW%	
Group					
1- Control (-ve)	25	0.039±0.001 ^a	28.54±1.04 ^a	0.88±0.05a	
2- Control (+ve)	16	0.029±0.001 ^c	14.06±0.93°	0.42±0.08f	
3-1ml Echinacea	18	0.036±0.001 ^{ab}	19.03±0.91b	0.52±0.08e	
4- 2ml Echinacea	21	0.035 ± 0.002^{ab}	21.76±2.22 ^b	0.61±0.11d	
5- 1ml Clove	18	0.036±0.003 ^{ab}	19.45±1.20 ^b	0.52±0.09e	
6- 2ml Clove	20	0.037±0.002 ^{ab}	22.07±1.74 ^b	0.64±0.08d	
7- 1ml E:C	22	0.036±0.001 ^{ab}	23.05±0.77 ^b	0.71±0.15c	
8- 2ml E:C	22	0.036±0.002 ^a	23.59±1.93 ^b	0.77±0.16b	

*Mean values are expressed as means ± SE.

*Mean values at the same column with the same superscript letters are not statistically significant at P<0.05.

***E:C= Echinacea: Clove.**

Results presented in **Table 2**, revealed that rats inflicted with immune deficiency diseases by cisplatin had significant (P < 0.05) reduction in serum immunogolobulin G (IgG) and immunogolobulin M (IgM) antibodies levels when compared with the negative control (-ve) group. Administration with *Echinacea* and Clove extracts and their mixture to IDD rats resulted in significant (P < 0.05) increases in serum IgG and IgM as compared to the positive control group. It was also observed that rats administrated with 2 ml mixture of *Echinacea* and Clove extracts recorded the best results for increasing IgG and IgM when compared to the negative control group.

Results in **Table 2**, are confirmed by **Nassef** *et al.*, (2018) who reported that cisplatin injection to rats caused a significant reduction in serum immunoglobulin (IgG, IgM). **Rehman** *et al.*, (1999) **Mahmoud** *et al.*, (2022), showed that *Echinacea* administration increased IgG and IgM production in rats with immune deficiency. The effects of immune activation by *Echinacea* were investigated by measuring total immunoglobulin (IgG, IgM). **Mishima** *et al.*, (2004) investigated the effects of immune activation by *Echinacea* by measuring T lymphocyte subsets in the peripheral blood of mice following whole-body irradiation

and reported that *Echinacea* activates macrophages to stimulate IFNgamma production in association with the secondary activation of T lymphocytes, resulting in decreases of IgG and IgM production. Also, the improvement of IgG and IgM may be due to that clove act as additional bonds with immunoglobulin molecules at the Fc receptors, which stimulated the immune response (Ahmed *et al.*, 2013).

Table 2: Effect of Echinacea and Clove Extracts on SerumImmunogolobulin G (IgG) and Immunogolobulin M(IgM) of Rats with Immune Deficiency Diseases

Parameter			
Group	IgG	IgM	
	(g /L)		
1- Control (-ve)	12.00±0.57 ^a	252.33±3.56 ^a	
2- Control (+ve)	5.46±0.88e	127.34±2.04 ^e	
3- 1ml Echinacea	6.90±0.62d	219.66±1.76 ^{cd}	
4- 2ml Echinacea	8.32±0.18c	231.66±3.35 ^{bc}	
5- 1ml Clove	6.89±0.16d	212.33±2.84 ^d	
6- 2ml Clove	8.46±0.32 ^c	233.66±4.25 ^{bc}	
7- 1ml E:C	9.80±0.21b	231.00±2.40 ^{bc}	
8- 2ml E:C	10.72±0.15b	235.52±1.15b	

*Mean values are expressed as means ± SE.

*Mean values at the same column with the same superscript letters are not statistically significant at P<0.05.

*E:C= Echinacea: Clove.

Results illustrated in **Table 3**, show that rats injected intraperitoneally with cisplatin had significant decreases in the serum levels of total protein, albumin and globulin when compared with the negative control group. Oral administration of Echinacea and Clove extracts (2 ml) and their mixture to IDD rats significantly normalized (P < 0.05) the levels of total protein, albumin and globulin in the serum. These findings are confirmed by Parameshappa et al., (2012) and Khalaf et al., (2019), who observed significant reduction in total protein, globulin and albumin concentration in rats administrated with cisplatin. Results also, are in agreement with that of Sadigh-Eteghad et al., (2011), who showed that treatment with Echinacea (500 mg /4 weeks) ameliorates the alteration in total protein and albumin. Concerning results of clove administration, are in agreement with Abozid and El-Saved (2013), who found that rats treated with clove extract increased plasma total protein and albumin compared with +ve group due to the polyphenolic compound and flavonoids that present in clove extract (Gulcin et al., 2004).

Deficiency Diseases					
Parameter	Total Protein	Albumin mg/dl	Globulin		
Group	mg/dl		mg/dl		
1- Control (-ve)	7.82±0.75a	4.85±0.61a	3.57±0.07a		
2- Control (+ve)	3.87±0.89d	2.15 ± 0.16^{d}	1.66 ± 0.12^{d}		
3- 1ml Echinacea	4.55±0.42cd	2.36±0.26 ^{cd}	1.89±0.12d		
4- 2ml Echinacea	5.10±0.27bc	3.24±0.15b	2.37±0.22c		
5- 1ml Clove	4.42±0.20cd	2.38±0.29cd	1.83±0.21d		
6- 2ml Clove	5.24±0.55bc	3.13±0.16bc	2.82±0.60b		
7- 1ml E:C	5.48±0.25b	3.32±0.83b	3.04±0.17b		
8- 2ml E:C	6.00±0.12b	3.38±0.63b	3.17±0.14b		

Table 3: Effect of Echinacea and Clove Extracts on Serum TotalProtein, Albumin and Globulin of Rats with ImmuneDeficiency Diseases

*Mean values are expressed as means ± SE.

*Mean values at the same column with the same superscript letters are not statistically significant at P<0.05.

***E:C= Echinacea: Clove.**

Results in **Table 4**, show that the positive control group had a significant increase (P < 0.05) in leukocytes and a significant decrease (P < 0.05) in levels of neutrophil, lymphocyte, monocytes and eosinophil as compared to the negative control group. On the other hand, rats that administrated with different levels of *Echinacea* & Clove extracts and their mixture had significant reduction in leukocytes and a significant elevation in neutrophil, lymphocyte, monocytes and eosinophil as compared to the positive control (+ve) group. The highest improvement was recorded in group that treated with the high level (2 ml 1E:1C) of combination of *Echinacea* and Clove extract.

In complementary of the present results, **Khalaf** *et al.*, (2019) found that administration of cisplatin to mature rats resulted in marked immunotoxic effects represented by leukopenia, lymphocytopenia and neutrophilia. On the other hand, **Markovic** *et al.*, (2011) reported that cisplatin increased the number of leukocytes due to the consequence of infection and inflammation. On the other hand, administration with *Echinacea* extract significantly reduced leucopenia induced by cisplatin which indicates that the extract could stimulate the haemopoetic system. This may be attributed to the contents of Echinacea as cichoricacid and echinacocide that stimulate bone marrow and the reformation of hematopoietic stem cells (Chopra and Goel 2002). Mishima *et al.*, (2004) reported that administration of Echinacea 360 mg/kg/day for 3weeks increases the number of leukocytes. This elevation is due to ability of polysaccharides and echinacocide to increase the number of

leukocytes. Doha et al., (2011) showed that Echinacea is involved in the modulation of immune response. Various phytoconstituents present in Echinacea, such as caffeic acid derivatives, alkamides, flavonoids, essential oils and polyacetylenes, are known to activate the non-specific cellular and humoral by increasing the production and activation of leukocytes, lymphocytes, monocytes and cytokines (Kim et al., 2002). These components also modulate the immune response by macrophage phagocytosis, pro-inflammatory cytokine production, activation of NK cell activity, enhancement of B cell response, increased T cell proliferation and elevated production of T cell cytokines (Chen et al., 2005; Thygesen et al., 2007 and Khalaf et al., (2019). Hence, it is plausible that *Echinacea* extract may confer immunoprotection through multifactorial immunomodulatory effects which could be achieved through chemical synergy of various bioactive constituents. On the other hand, Dibazar et al., (2015) and Kmiec et al., (2017), revealed that clove extract containing eugenol, β -caryophyllene, caryophyllene oxide and α -humulene can increase the proliferation of activity of lymphocytes, lymphoblasts and reactive oxygen intennediate secretion of macrophages. Infected mice will increase IFN- γ levels because there are immunogens that activate the immune system.

Table 4: Effect of Echinacea and Clove Extracts on Leukocytes,
Neutrophil, Lymphocyte, Monocytes, Eosinophil, and
Basophil Count of Rats with Immune Deficiency Diseases

Parameter	Leukocytes	Neutrophil	Lymphocyte	Monocytes	Eosinophil	Basophil
Group	$(x \ 10^{3}/ul)$	(x 10 ³ /ul)				
1- Control (-ve)	5.30±0.57e	5.41±0.12 ^a	4.83±0.19 ^a	$1.84{\pm}0.07^{a}$	1.37±0.02a	1.50±0.05a
2- Control (+ve)	12.03±0.24 ^a	0.79±0.02 ^e	1.01±0.05f	0.35 ± 0.02^{d}	0.30±0.03f	0.19±0.01e
3- 1ml Echinacea	11.22±0.18b	1.52 ± 0.19^{d}	1.87±0.05e	0.49±0.03 ^{cd}	0.55±0.02de	0.33±0.01de
4- 2ml Echinacea	8.40±0.15 ^c	2.83±0.09°	2.80±0.09d	0.56±0.01°	0.71±0.01cd	0.47±0.03d
5- 1ml Clove	10.88±0.34 ^b	1.73 ± 0.09^{d}	1.69±0.09e	0.46±0.01 ^{cd}	0.49±0.01e	0.29±0.01de
6- 2ml Clove	7.99±0.26°	2.71±0.14 ^c	2.91±0.03cd	0.63±0.03°	0.80±0.01bc	0.51±0.01d
7- 1ml E:C	6.73±0.15 ^d	3.03±.04 ^c	3.10±0.06c	0.58±0.04 ^c	0.79±0.04bc	0.88±0.01c
8- 2ml E:C	6.33±0.12 ^d	4.02 ± 0.10^{b}	4.18±0.10b	0.94±0.03 ^b	0.94±0.01b	1.16±0.09b

*Mean values are expressed as means ± SE.

*Mean values at the same column with the same superscript letters are not statistically significant at P<0.05.

*E:C= Echinacea: Clove.

CONCLUSION

The findings of this study demonstrated that Echinacea and clove extract is a promising immunomodulatory agent with a potent therapeutic value in stimulating the suppressed immune response.

REFERENCES

- Abozid, M. and S. El-Sayed (2013): Antioxidant and protective effect of clove extracts and clove essential oil on hydrogen peroxide treated rats. Int. J. Chem. Technol., Res., 5: 1477-1485.
- Agbaje, E. ; A. Adeneye and A. Daramola (2009): Biochemical and toxicological studies of aqueous extract of *Syzigium aromaticum* (L.) Merr. & Perry (Myrtaceae) in rodents. AJTCAM, 6: 241–254.
- Ahmed, S.; M. Hossain; G. Kim; J. Hwang; H. Ji and C. Yang (2013): Effects of resveratrol and essential oils on growth performance immunity, digestibility and fecal microbial shedding in challenged piglets. Asian-Australas J. Animal Sci., 26: 683- 690.
- Ali, E. (2008): Protective effects of *Echinacea* on cyproterone acetate induced liver damage in male rats. Pakistan J. Biol. Sci., 11: 2464–2471.
- Amelia, B.; E. Saepudin ; A. Cahyana ; D. Rahayu ; A. Sulistyoningrum and J. Haib (2017): GC-MS analysis of clove (*Syzygium aromaticum*) bud essential oil from Java and Manado, AIP Conference Proceedings, AIP Publishing LLC, 030082.
- Armitage, G.Y. and W.G. Berry (1987): Statistical methods 7th Ed. Ames., Iowa State University. Press.39-63.
- Bachiega, T. ; J. de Sousa ; J. Bastos and J. Sforcin (2012): Clove and eugenol in non-cytotoxic concentrations exert immunomodulatory/ anti-inflammatory action on cytokine production by murine macrophages. J. Pharm. Pharmacol. 64:610–616.
- Burtis, C. and E. Ashwood (1999): Tietz Textbook of Clinical Chemistry. 3rd ed. Philadelphia: W.B. Saunders, 1999: 1840, 1841, 1844, 1845; 1799; 1834-5 Textbook of Clinical Chemistry, 3rd . AACC.
- Chaieb, K. ; H. Hajlaoui and T. Zamantar (2007): The chemical composition and biological activity of clove essential oil, *Eugenia caryophyllata (Syzugium aromaticum* L *Myrtaceae*): A short review. Phytother. Res., 21:501–506.
- Chapman, D.G.; R. Gastilla and J.A. Campbell (1959): Evaluation of protein in foods: 1- A Method for the determination of protein efficiency ratio. Can. J. Biochem. Phys., 37:679-686.
- Chen, Y.; T. Fu; T. Tao; I. Yang; Y. Chang and L. Kim (2005): Macrophage activating effect of new alkamides from the roots of Echinacea species. J. Nat. Prod., 68: 773–776.
- Chiou, S.; J. Sung; P. Huang and S. Lin (2017): antioxidant, antidiabetic, and antihypertensive properties of echinacea purpurea flower extract and caffeic acid derivatives using *In Vitro* models. J. Med. Food, 20:171-179.

- Chopra, A.K. and R.K. Goel (2002): A modal pushover analysis procedure for estimating seismic demands for buildings. Earthquake Engng. Struct., 31:561–582
- Cortés-Rojas, D. ; C. de Souza and W. Oliveira (2014): Clove (Syzygium aromaticum): A precious spice. Asian Pac. J. Trop. Biomed., 4: 90–96.
- **Dasari, S. and P. Tchounwou (2014):** Cisplatin in cancer therapy: molecular mechanisms of action. Eur. J. Pharmacol., 740: 364–378.
- **Dibazar, S.; S. Fateh and S. Daneshmandi (2015):** Immunomodulatory effects of clove (*Syzygium aromaticum*) constituents on macrophages: *In Vitro* evaluations of aqueous and ethanolic components. J. Immunotoxicol., 12(2): 124-131.
- **Doha, E.H.; A.H. Bhrawy and S.S. Ezz-Eldien (2011):** A Chebyshev spectral method based on operational matrix for initial and boundary value problems of fractional order. Computers & Mathematics with Applications, 62 (5):2364-2373.
- El-Sherbiny, E. ; H. Osman and M. Taha (2021): Effectiveness of *Echinacea purpurea* extract on immune deficiency induced by azathioprine in male albino rats. Biosci. J., 37: 1-12.
- Grespan, R. ; M. Paludo and H. Lemos (2012): Anti-arthritic effect of eugenol on collagen-induced arthritis. Experimental model. Biol. Pharm. Bul. 35: 1818–1820.
- Gulcin, I.; I. Sat; S. Beydemir; M. Elmastas and O. Kufrevioglu (2004): Comparison of antioxidant activity of clove (*Eugenia* caryophylata Thunb) buds and lavender (*Lavandula stoechas* L.). Food Chem., 87: 393–400.
- Hasaaan, I.; S. Chibber and I. Naseem (2010): Ameliorative effect of riboflavin on the cisplatin induced nephrotoxicity and hepatotoxicity under photoillumination. Food Chem. Toxicol., 48: 2050–2058.
- Khalaf, A.; S. Hussein; A. Tohamy; S. Marouf; H. Dawood; A. Zaki and A. Bishayee (2019): Protective effect of Echinacea purpurea (Immulant) against cisplatin-induced immunotoxicity in rats, DARU J. Pharmaceutical Sci., Daru, 27: 233–241.
- Khattab, H.; S. Abounasef and H. Bakheet (2019): The biological and hematological effects of *Echinacea purpurea* L. roots extract in the immunocompromised rats with cyclosporine. J. Microscopy and Ultrastructure, 7: 65-71.
- Kim, L. ; R. Waters and P. Burkholder (2002): Immunological activity of larch arabinogalactan and Echinacea: A preliminary, randomized, double-blind, placebo-controlled trial. Altern Med. Rev., 7: 138–149.

- Kmiec, Z. ; M. Cyman and T. Slebioda (2017): Cells of the innate and adaptive immunity and their interactions in inflammatory bowel disease. Adv. Med. Sci., 62:1-16.
- Lalone, C. ; K. Hammer ; L. Wu ; J. Bae ; N. Leyva ; Y. Liu ; A. Solco ; G. Kraus ; P. Murphy ; E. Wurtele ; O. Kim ; M. Widrlechner, and D. Birt (2007): Echinacea species and alkamides inhibit prostaglandin E(2) production in RAW264.7 mouse macrophage cells. J. Agric. and Food Chem., 55: 7314-7322.
- Lin, M. ; J. Ko ; T. Liu ; P. Chao and C. Chu-Chyn Ou (2018): Protective effect of d-methionine on body weight loss, anorexia, and nephrotoxicity in cisplatin-induced chronic toxicity in rats. Integr. Cancer Ther., 17: 813–824.
- Mahmoud, A. ; M. Abbas and H. AbdElmonem (2022): The Antioxidant effects of cerium oxide nanoparticles and *Echinacea purpurea* against lead-induced immunosuppression in male albino rats. The Egyptian J. Hospital Med., 89: 6106-6114.
- Markovic, S. ; J. Z'izic ; D. Djacic ; A. Obradovic ; D. Cvetkovic ; N. Ordevic ; B. Ognjanovic and A. Stajn (2011): Alteration of oxidative stress parameters in red blood cells of rats after chronic *In Vivo* treatment with cisplatin and selenium. Arch. Biol. Sci., 63: 991–999.
- Mishima, S.; K. Saito ; H. Maruyama ; M. Inoue ; T. Yamashita ; T. Ishida and Y. Gu (2004): Antioxidant and Immuno-Enhancing Effects of Echinacea purpurea. Biol. and Pharmaceutical Bull., 27:1004-1009.
- Nagoba, B. and M. Davane (2018): Natural Immunomodulators. J. Immunology and Microbiol., 2: 1:2.
- Nassef, N. ; M. El-Melegy ; E. Beshay ; D. Al-Sharaky and T. Al-Attar (2018): Trypanocidal effects of cisplatin alone and in combination with *Nigella sativa* oil on experimentally infected mice with *Trypanosoma evansi*. Iran J. Parasitol., 13: 89–99.
- Ochei, J. and A. Kolhatkar (2008): Medical Laboratory Sciences; Theory and Practice. Tata McGraw-Hill Publishing Co. Ltd. New Delhi; 321-324.
- Parameshappa, B.; M. Basha ; S. Sen ; R. Chakraborty ; G. Kumar ; G. Sagar ; L. Sowmya ; K. Raju; K. Sesh and A. Lakshmi (2012): Acetaminophen induced nephrotoxicity in rats: Protective role of Cardiospermum halicacabum. Pharm. Biol. 50:247–253.
- Pearse, G. ; A. Pietersma ; J. Cunliffe ; J. Foster ; J. Turton ; N. Derbyshire and K. Randall (2009): Time-course study of the immunotoxic effects of the anticancer drug chlorambucil in the rat. Toxicol. Pathol., 37: 887–901.

- Reeves, P.; F. Nielsen and G. Fahmy (1993): AIN-93. Purified diets for laboratory rodents: Final reports of the American Institute of Nutrition adhoe wriling committee of reformulation of the AIN-76 A Rodent Diet. J. Nutr., 123: 1939-1951.
- Rehman, J. ; M. Dillow ; J.S. Carter ; A. Chou ; B. Le and A. Maisel (1999): Increased production of antigen-specific immunoglobulins G and M following *In Vivo* treatment with the medicinal plants *Echinacea angustifolia* and hydrastiscanadensis. Immunol. Letters, 68: 391-5.
- Sadigh-Eteghad, S. ; H. Khavat-Nuri ; N. Abadi ; S. Ghavami ; M. Golabi and D. Shanebandi (2011): Synergetic effects of oral administration of *Levamisole* and *Echinacea purpurea* on immune response in wistar rat. Res. Vet. Sci., 91: 82-85.
- Sharifi-Rad, M. ; D. Mnayer ; M. Morais-Braga ; J. Carneiro ; C. Bezerra ; H. Coutinho ; B. Salehi ; M. Martorell ; M. Contreras ; A. Soltani-Nejad ; Y. Uribe ; Z. Yousaf ; M. Iriti and J. Sharifi-Rad (2018): Echinacea plants as antioxidant and antibacterial agents: From traditional medicine to biotechnological applications. Phytherapy Res., 32: 1653–1663.
- Thygesen, L. ; J. Thulin ; A. Mortensen ; L. Skibsted and P. Molgaard (2007): Antioxidant activity of cichoric acid and alkamides from *Echinacea purpurea*, alone and in combination. Food Chem., 101: 74–81.
- Tumeh, P.; C. Harview; J. Yearley; I. Shintaku; E. Taylor; L. Robert
 ; B. Chmielowski; M. Spasic; G. Henry; V. Ciobanu; A. West
 ; M. Carmona; C. Kivork; E. Seja; G. Cherry; A. Gutierrez;
 T. Grogan; C. Mateus; G. Tomasic; J. Glaspy; R. Emerson;
 H. Robins; R. Pierce; D. Elashoff; C. Robert and A. Ribas
 (2014): PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature, 515: 568-571.
- Wang, Y.; L. Chen; G. Huang; D. He; J. He and W. Xu (2013): Klotho sensitizes human lung cancer cell line to cisplatin via PI3k/Akt pathway. PLoS One, 8: 573–591
- Yogalakshmi, B. ; P. Viswanathan and C. Anuradha (2010): Investigation of anti-oxidant, anti-inflammatory, and DNAprotective properties of eugenol in thioacetamide-induced liver injury in rats. Toxicol., 268:204–212.
- Young, D. (1995): Effect of drugs on clinical lab Tests, 4 th ed AACC press.
- Zhang, D. ; K. Wu ; X. Zhang and S. Deng (2020): Peng, B. In silico screening of Chinese herbal medicines with the potential to directly inhibit 2019 novel coronavirus. J. Integr. Med., 18, 152–158.
- Zhou, Z. ; Y. Zhao ; S. Chen ; G. Cui ; W. Fu ; S. Li ; X. Lin and H. Hu (2022): Cisplatin promotes the efficacy of immune checkpoint

inhibitor therapy by inducing ferroptosis and activating neutrophils. Front Pharmacol., 13: 1-16.

- Zhu, X. ; X. Jiang ; A. Li ; Z. Zhao and S. Li (2017): S-Allylmercaptocysteine attenuates cisplatin-induced nephrotoxicity through suppression of apoptosis, oxidative stress, and inflammation. Nutrients, 9: 166.
- Ziva, J. and P. Pannall (1984): Clinical Chemistry In Diagnosis And Treatment. Publ. L. Loyd-Luke (Medical Books), Londo, 348-352.

تأثير المستخلص المائي للإكنيشيا والقرنفل على نقص المناعة المحدث بواسطة

السيسبلاتين في الفئران

أسماء محمود عبدالحافظ ، أمنية جلال رفعت ، هاني جابر المصري ، آلاء أسامة أبو ريه

قسم التغذية وعلوم الأطعمة - كلية الاقتصاد المنزلي - جامعة حلوان

هدفت الدراسة الحالية إلى معرفة تأثير المستخلص المائي للإكنيشيا والقرنفل في رفع الاستجابة المناعية في الفئران المصابة بأمراض نقص المناعة. تم تقسيم 48 من ذكور الجرذان البيضاء بشكل عشوائي إلى 8 مجموعات متساوية (ن = 6). المجموعة الأولى كانت الضابطة السالبة بينما تم حقن المجموعات السبع الأخرى بواسطة سيسبلاتين. احتفظت المجموعة 2 كمجموعة ضابطة موجبة للسمية المناعية. تلقت المجموعتان 3 ، 4 مسنخلص الإكنيشيا بجرعة 1 و 2 مل ، في حين أعطيت المجموعتان 5 و 6 على التوالي 1 ، 2 مل من مستخلص القرنفل. تلقت المجموعتان 7 ، 8 خليط من مستخلص الإكنيشيا والقرنفل بجرعة 1و 2 مل، على التوالي ، لمدة 8 أسابيع. تشير نتائج وزن الجسم ، وزن الطحال ، الجلوبيولين المناعي (IgM , IgG) ، البروتين الكلى ، الألبومين ، الجلوبيولين انخفضت بدرجه معنويه. بينما زادت الكريات البيض بشكل ملحوظ بواسطة السيسبلاتين في المجموعة الضابطة الموجبة مقارنة بالمجموعة الضابطة السالبة. من ناحية أخرى ، فإن إعطاء مستخلص الإكنيشيا والقريفل وخليطيهما قد خفف من هذه الآثار الضارة وخفف بشكل ملحوظ التغيرات الكيميائية الحيوية التي تسببها حقن سيسبلاتين. في الختام ، يحفز مستخلص الإكنيشيا والقرنفل الجهاز المناعي للفئران التي تعانى من نقص المناعة المُحدث بواسطة السيسبلاتين. أوصت هذه الدراسة بزيادة استهلاك الإكنيشيا والقرنفل في النظام الغذائي، فقد يؤدي هذا إلى رفع الاستجابة المناعية لدى المريض المصاب بإضطرابات مناعية.